



# Science in School

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## Plant solar power: unlocking the secrets of photosynthesis with X-ray free-electron lasers

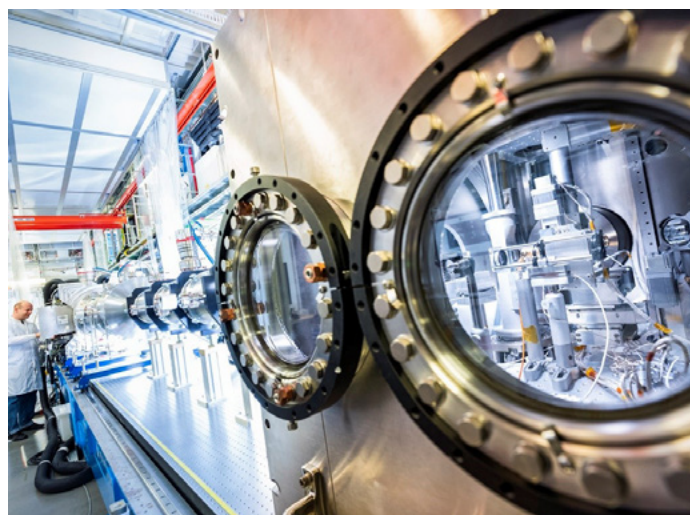
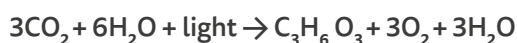
Rosemary Wilson

Capturing the moment: to use artificial photosynthesis to produce clean energy, we need to better understand it first. Learn about how X-ray free-electron lasers can help.

Photosynthesis is a remarkable process that enables plants to effectively convert the sun's energy into chemical energy. It uses molecules abundant in the atmosphere to produce energy that can be stored, and even by-products of photosynthesis are beneficial for other organisms. The latest solar-energy technologies do not come close to fulfilling these criteria, so imagine if we could mimic photosynthesis to meet our own energy needs. Before we start, we need to understand precisely how photosynthesis works. But photosynthesis is a complex process, involving large assemblies of proteins, and occurs on an unfathomably short timescale, so how can we get a true picture of what is going on?

### What do we know about photosynthesis?

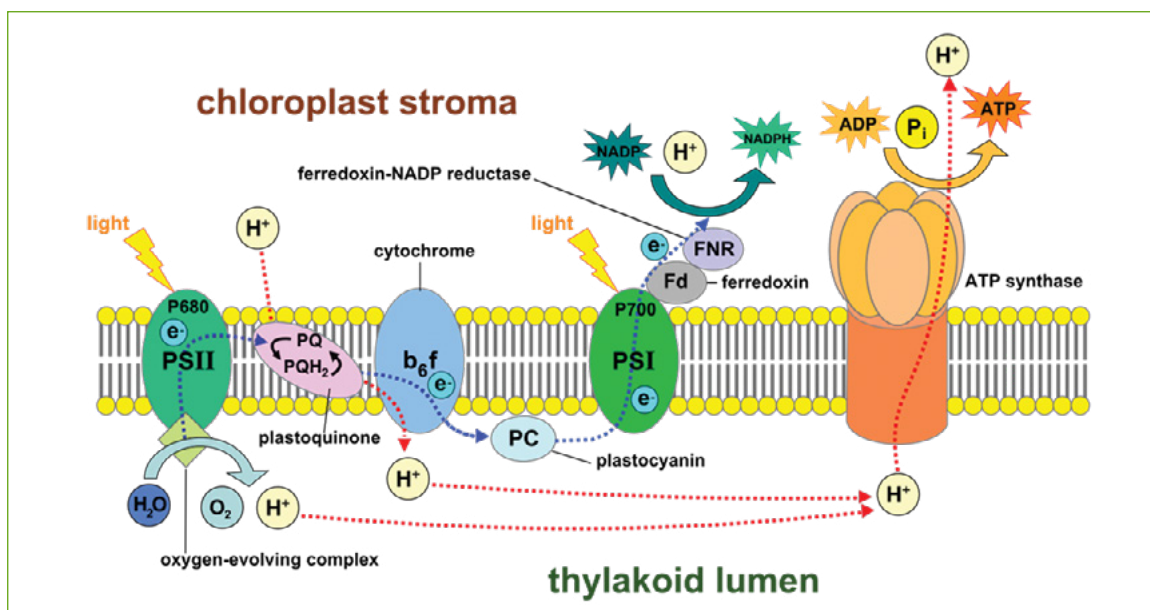
Photosynthesis is the process by which green plants, cyanobacteria, and algae capture and use light energy to convert carbon dioxide and water into chemical energy in the form of sugars. It is summed up in the following equation:



The serial femtosecond crystallography instrument at the European XFEL.

*European XFEL*

The chloroplast, a cell organelle unique to photosynthesizing organisms, is the solar-power centre of the plant. It contains two types of large, light-sensitive, protein machines called photosystems – photosystem I (PSI) and photosystem II (PSII).

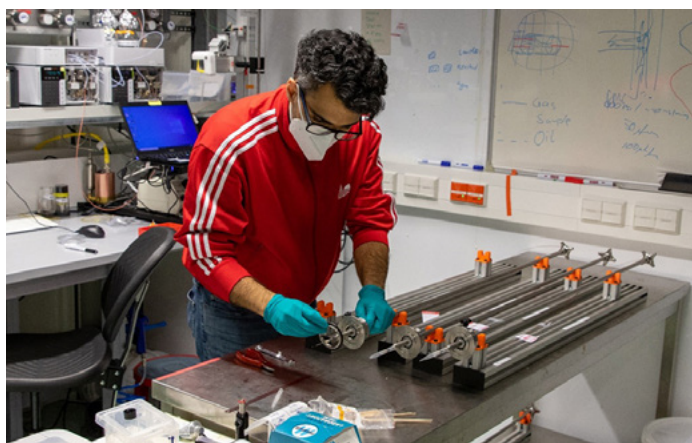


Arrangement of the photosystems on the thylakoid membrane, including details of the photosynthetic pathways.

Somepics/ [Wikimedia](https://commons.wikimedia.org/wiki/File:Thylakoid_membrane.jpg), [CC BY-SA 4.0](https://creativecommons.org/licenses/by-sa/4.0/).

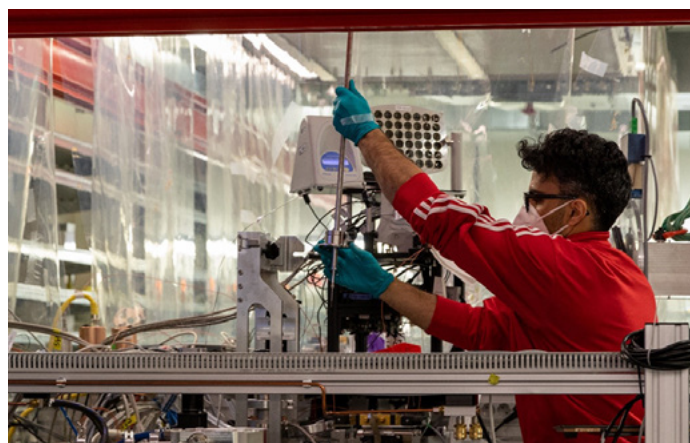
The photosystems, located in the thylakoid membrane, capture photons of light and move them to chlorophyll molecules, which absorb the photons, become excited, and release electrons. The electrons move through an electron-transfer chain from one photosystem to the other, before they are used to create energy (by converting nicotinamide adenine dinucleotide phosphate (NADP<sup>+</sup>) into the reduced form, NADPH) for the Calvin cycle to produce sugars from carbon dioxide. Water is also a crucial component; PSII splits water molecules to release oxygen molecules and hydrogen ions. These ions build up to create a charge difference across the thylakoid membrane, and then move across the membrane via adenosine triphosphate (ATP) synthase, creating energy in the form of ATP.

If we want to build our own solar-energy cell, we need to know more about the 3D atomic structures of PSI and PSII and what goes on inside them. While researchers have studied photosynthesis for decades, it has been hard to pinpoint some of the details. Scientists can use high-powered X-ray beams [to create models of a protein's structure](#), but first the proteins must be crystallized. Large protein crystals work best, but growing them is not always easy. Photosystems are huge protein complexes – PSI can consist of around 26 proteins, 288 chlorophylls, and 380 other molecules – and large proteins are especially difficult to crystallize. Membrane proteins, like PSI and PSII, are even more difficult to crystallise. At best, large proteins and membrane proteins produce tiny crystals, which need long exposure times, so the powerful X-ray radiation could destroy the proteins before we get our image.



Preparing the jet nozzles in the lab: a jet of tiny crystals will be passed through this into the X-ray beam.

European XFEL



Mounting the jet nozzle into the instrument.

European XFEL

## How can X-ray free-electron lasers (XFELs) help?

### European XFEL

The European XFEL is an X-ray free-electron laser that generates extremely short and bright X-ray flashes at a rate of about 27 000 times per second. These X-ray flashes are around a billion times brighter than those produced at synchrotrons – another type of X-ray light source. The European XFEL is located in underground tunnels that cover a distance of 3.4 km, from the Deutsches Elektronen-Synchrotron (DESY) campus in Hamburg to the neighbouring town of Schenefeld. To generate X-ray light, electrons are first accelerated to almost the speed of light. After they have raced through the 1.7-km-long accelerator, the electrons pass into structures called undulators. These consist of a series of alternating magnets that force the electrons to go on a tight zigzag course. When electrons are sent on a curved path by strong magnets like this, they emit laser-like X-ray light. Scientists from across the world come to the European XFEL to use these X-ray flashes for their research.



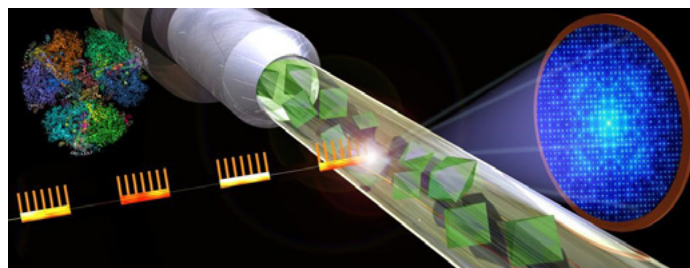
Inside the European XFEL tunnel. The yellow tubes are where electrons are accelerated.

European XFEL

### Detailed images of large proteins

Light produced at the European XFEL is extremely bright and is generated in very short bursts that arrive in quick succession, which means that it can be used to overcome some of the challenges associated with the imaging of large membrane proteins, such as PSI and PSII. At the European XFEL, it doesn't matter if the crystals are small. In fact, a method called serial femtosecond crystallography has been optimized to use microcrystals. First, a jet of tiny crystals is fired into the path of the X-ray beam. The crystals are then

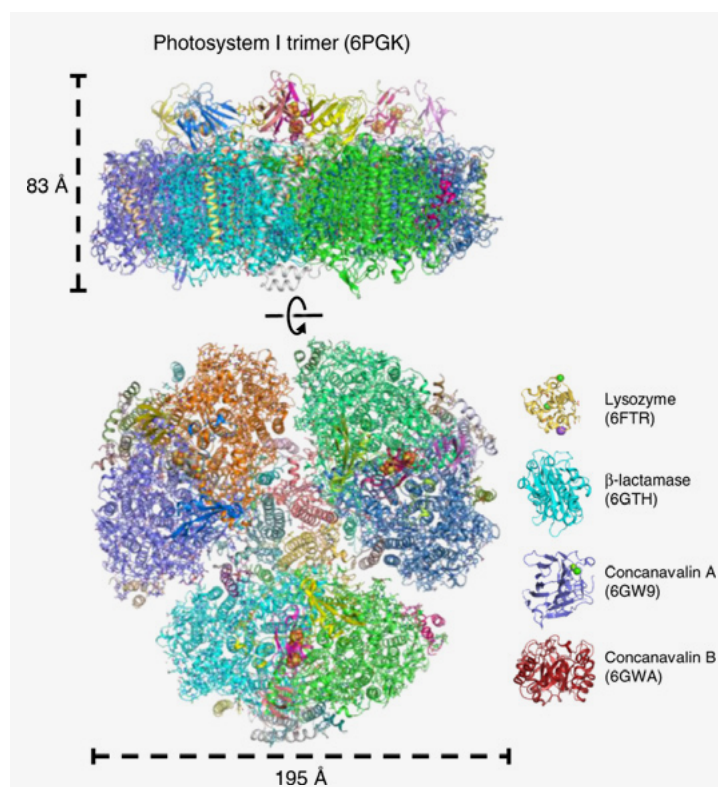
blasted by a flash of X-ray light, creating an image of each protein before the powerful radiation destroys it.



Basic design of a serial femtosecond crystallography experiment at the European XFEL. X-ray bursts (symbolized by the golden combs coming in from the left) strike crystallized samples, resulting in diffraction patterns that can be reassembled into detailed images.

Shireen Dooling/ASU

The individual images are put together to create a detailed composite image. Scientists have already used the fast rate of flashes produced by the European XFEL in this way, and XFELs have been used to create the most detailed images we have of PSI.<sup>[1,2]</sup>



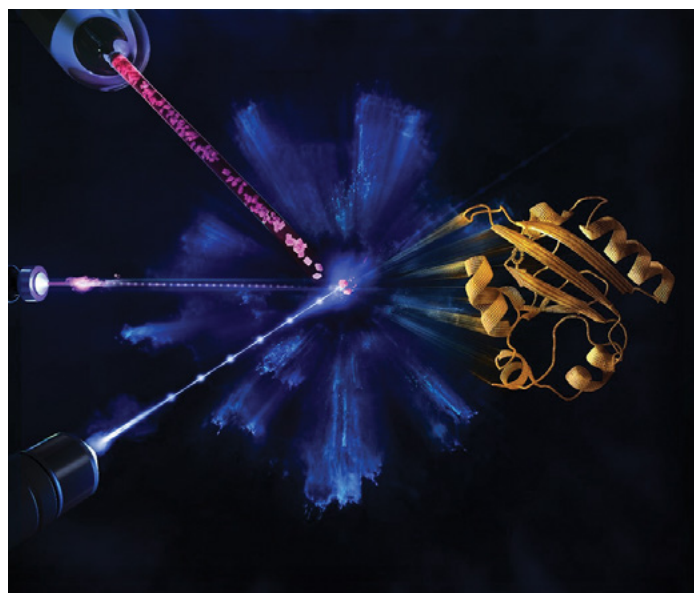
Structure of PSI based on data collected at the European XFEL during one of the first beam-times in September 2017. Single proteins are shown on the right for comparison. The side-on view at the top shows details of the antennae structures above the main body of the structure; these antennae capture light particles and funnel them down to the reaction centre in the middle of the molecule. The size of the molecules is measured in Ångstrom; 1 Å is roughly the size of one atom or 0.1 nanometres. PSI is a trimer, meaning that it is built from three subunits, known as monomers.

From Ref. [1], [CC BY 4.0](#).

## Molecules in action

However, even a high-quality image is just a single moment captured in time. It allows us to capture a detailed before and after shot, but what we really want is to be able to see these proteins in action, to understand what the intermediate steps are. Only then can we begin to understand, for example, how PSII splits water. Photosynthesis is, however, an extremely fast process. Energy transfer between different parts of the photosystem takes around 500–1000 femtoseconds. A femtosecond is one quadrillionth of a second or  $1 \times 10^{-15}$  s. For comparison, a mosquito can beat its wings up to 1000 times a second, so each wingbeat takes 100 000 000 000 ( $1 \times 10^{11}$ ) femtoseconds. The entire process of photosynthesis is over before a mosquito flaps its wings.

A photographer taking images of a football match chooses the shutter speed of their camera to match the speed of the action, to ensure they can take a series of sharp images in quick succession. If they get enough good images, the story of the match can be pieced together later. Likewise, to document the progression of photosynthesis, we need to have a 'camera' that is fast enough to capture that movement. The X-ray flashes generated at the European XFEL are so short that scientists can take sequences of images of processes occurring on the femtosecond timescale and stitch them together into a 'molecular movie'.



Artist's impression of a serial femtosecond crystallography experiment to create images of a light-sensitive protein. A stream of crystalline proteins is first hit by an optical laser before being struck by the X-ray laser. Information recorded about the arrangement of atoms in the protein is used to reconstruct a model of the structure of the protein.

European XFEL / Blue Clay Studios



Scientists monitoring the experiment from the control room.  
European XFEL

With light-sensitive proteins, such as PSI and PSII, the reaction can be triggered by exposing the protein crystals to a light pulse. X-ray flashes are then timed to strike crystals at a series of timeframes throughout the reaction, so scientists can 'take' images of different stages of the process, as it unfolds.

At the European XFEL, scientists have tested this experimental setup using another light-sensitive protein and will now use the setup to make molecular movies of PSI and PSII.<sup>[3,4]</sup>

## Outlook for artificial photosynthesis

Unlike plants, our mobility and industrial systems aren't powered by sugar, but imitating parts of the photosynthetic process would allow us to produce useful products. We could split water to create hydrogen, which could be used to fly planes or make steel, or we could use carbon dioxide to create fuels, such as methanol, for vehicles or feed these fuels into chemical processes. Scientists can already mimic some of these processes in the lab. With our latest understanding of physics paired with high-precision engineering, we can begin to reveal the secrets of this amazing natural process and achieve new efficiencies for a sustainable future. <<

## References

1. Gisriel C et al. (2019) [Membrane protein megahertz crystallography at the European XFEL](#). *Nature Communications* **10**:5021. doi: 10.1038/s41467-019-12955-3
2. Using the European XFEL to shed light on photosynthesis: [https://www.xfel.eu/news\\_and\\_events/news/index\\_eng.html?openDirectAnchor=1733&two\\_columns=0](https://www.xfel.eu/news_and_events/news/index_eng.html?openDirectAnchor=1733&two_columns=0)

3. First molecular movies at the European XFEL: [https://www.xfel.eu/news\\_and\\_events/news/index\\_eng.html?openDirectAnchor=1736&two\\_columns=0](https://www.xfel.eu/news_and_events/news/index_eng.html?openDirectAnchor=1736&two_columns=0)
4. Pandey S et al. (2020) [Time-resolved serial femtosecond crystallography at the European XFEL](#). *Nature Methods* **17**: 73–78. doi: 10.1038/s41592-019-0628-z.

## Resources

- Learn more about solving protein structures: Heber S (2021) [From gaming to cutting-edge biology: AI and the protein folding problem](#). *Science in School* **52**.
- Learn about the PDB Art project and how you can use it in your class:
  - Gupta D, Armstrong D (2021) [Introducing students to the beauty of biomolecules](#). *Science in School* **53**.
  - Gupta D, Armstrong D (2021) [Bringing the beauty of proteins to the classroom: the PDB Art Project](#). *Science in School* **54**.
- Read about a classroom activity to discover the pigments that play a role in photosynthesis: Tarragó-Celada J, Fernández Novell JM (2019) [Colour, chlorophyll and chromatography](#). *Science in School* **47**:41–45.
- Use plants to investigate the relationship between green pigment and leaf growth: Patil K, Singh G, Haydock K (2015) [Do leaves need chlorophyll for growth?](#) *Science in School* **33**:47–51.
- Read an article on the development of [time-resolved crystallography](#), including a nice [infographic](#) that explains the process.
- Watch a video on [artificial leaf technology](#).
- If we could [watch trees in infrared](#), what colour would they be?
- Watch a video on common preconceptions of where [the mass of trees comes from](#).
- Take [a closer look at photosynthesis](#).
- Watch a video on [why plants are green](#) and not black.

## AUTHOR BIOGRAPHY

**Rosemary Wilson** is a science communicator and writer based in Germany. She currently works with the communications group at the European XFEL, before which she worked at the European Molecular Biology Laboratory (EMBL) in Hamburg. A long time ago, she did her PhD in plant molecular phylogeny, studying the molecular relationships within a group of liverworts.



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